# **Plants and Ecosystem**

ISSN: 2789-7370 (Online) Volume- 01 (2021)

**REVIEW ARTICLE** 

https://doi.org/10.54479/pae.v1i01.6666

# A Mini Review on Microalgae Biomass Production: Recent Progress in Cultivation Systems

ISMAIL RASEL | SAYDUL KARIM

Department of Biotechnology, South Asian University, New Delhi-110021, India

## Article History

Received: 05 August 2021 Revised: 21 September 2021 Accepted: 25 September 2021 Published: 10 October 2021

Corresponding Author Saydul Karim Contact: sdkbcs10@gmail.com **ABSTRACT:** Single-celled photosynthetic microscopic algae have numerous significances for human (microalgae) wellbeing. From medicine to fuel industry these tiny organisms have tremendous potentials and in future they would be game changer to mitigate global warming and environmental pollutions. The current production cost is a matter of consideration during its applications. Scientists all over the world are trying to reduce the production costs as well as to develop new or improve the existing culturing methodologies and techniques. Open pond and closed pond (PBRs) culturing system are two most prominent ways to culture microalgae. Open pond culturing techniques for microalgae have several advantages over closed pond system such as low operational costs and easy to make. However, the main drawback of this system is contamination by other microorganisms, which is possible to control in closed pond system. Among the many types of PBRs systems, the advanced tubular PBRs presently considered as more useful than open pond culture system. To meet the minimum production costs, more research is needed on both the culturing systems.

Keywords: Microalgae, Open Pond, PBRs, Algae biomass.

© The Author(s)-2021

A group of single-celled microscopic autotrophs are generally called microalgae in biology, which commonly produces atmospheric oxygen via photosynthesis using  $CO_2$  and water in the presence of sun light (1). They are considered as the most primitive form of plants because of their very simple cellular structures. They usually grow in suspension of a surface water of any aquatic environments and can double every few hours during their exponential growth time (2-4). Their motile and non-motile features make them versatile, and they can consume organic material for their energy requirements (5,6). Within the cell, they can synthesize all essential

amino acids, and can possesses high levels of proteins, carbohydrates as well as lipids. Algal species containing high protein contents could be used as an ideal source of functional foods, food additives and nutraceuticals precursor in commercial industries (7,9). Besides, it is also used as human foods, food supplements, pharmaceuticals, cosmetic industry, immune-modulating, anti-cancer products (10-12). Furthermore, C- sequestrations to abate global warming, bioremediation of heavy metals from wastewater, biotransformation, and treatment of sewage and municipal wastes are also considered as the potential usages of microalgae nowadays (13).

CC II

This is an Open Access (OA) article under a Creative Commons license: Attribution 4.0 International (CC-BY). No financial, legal or technical barriers to access this article. To see the terms and conditions- (*http://creativecommons.org/ficenses/by/4.0/*).

The worldwide need for energy usage is surprisingly increasing day by day. It is estimated that the demands will be much more than 85% by 2040 (14). Currently, most the energy demands in the globe are met by fossil fuel, although the resources which reducing gradually. Thus, the additional sources of renewable energy, particularly biofuels, must be considered for sustainable energy management (15). At present, plants and plants parts have been used as biofuel feedstock in many parts of the world with some limitations that pose new challenges for humankind. For example, the amount of arable lands is reducing which would contribute to food crises in near future. Microalgae have been considered as a suitable source for biofuels production due to their higher growth rates, higher photosynthetic efficiency as well as higher biomass productivity, as compared to other terrestrial plants or plants parts (16,17). According to a report, it is estimated that the produced algal oil per acre is almost 30 times higher than oil crops (15). Although microalgae biomass is regarded as the most suitable renewable feedstock for high-energy production, its pilot-scale economic utilization is still challenging due to its production costs along with other associated limitations. As only a small land area can support to produce biomass several times higher than currently used energy crops, microalgae still now attractive for bioenergy production purposes and regarded as a best competitor in this field for future utilization (18).

However, presently microalgae can be cultivated adopting different techniques and methodologies to make an expectable amount of biomass. Open pond systems, closed systems, and hybrid systems are the most prominent cultivating systems (19,20). However, open pond system is very easy to make and possible to install for pilot scale production contamination by purposes, although other unwanted microorganisms is a major disadvantage of open pond culturing system. A closed system, also called a photobioreactor (PBR), is now widely studied due to the facility of controlled growth and less contamination. It gives a higher ratio of biomass to substrate conversion with economic efficiency than an open system (21). However, the high cost of construction and maintenance of PBR systems restricted the use of the system mostly for research purposes till now (22). Construction of PBR systems, formulation of the growth medium, and maintaining the turbulent flow continuously is also a very highcost process that outweighs its other advantages (23). This high cost can be checked by the use some low-cost materials as PBRs unit and uses the wastewater or industrial surplus products as a growth medium and also possible to use an energyefficient pumps with the system (24). Commercial application of microalgae is still a dream.

Moreover, microalgae have the huge scopes to use in biotechnology industry targeting different demands in the future (25). As production cost is the major problem in microalgae biomass industry, there has been seen many improvements achieved during the last decades by the hard efforts of scientists (26-30). They are trying to develop new strategies to produce microalgae with minimum costs, which will be commercially viable to cultivate and to apply. Therefore, the main aim of the review was to study the recent findings developed by researchers under different type's affordable culturing techniques to produce microalgae biomass as cheap as possible.

# MATERIALS AND METHODS

To collect the information, different open access journals were scrutinized to find out recent research progress on microalgae cultivation. Besides, other paid articles also were downloaded with topic relevance. Most of the keywords that wrote in search engine are- microalgae culture, recent progress in microalgae culture, open pond culture, open pond culture for microalgae, closed cultivation of microalgae, photobioreactors, low cost culturing, and cost reduction approaches, so on. The collected articles were shorted based on the publishing year. Then the collected articles were arranged according to two types of major cultivation- open pond and photobioreactors (PBRs) systems.

# MICROALGAE CULTURING TECHNIQUES

Various types of cultivation techniques for algae biomass productions are being practiced for different purposes and based on algae types. Nevertheless, most of them are mainly based either on open ponds or closed pond culturing system. Moreover, hybrid type is also getting attention for coast lowering approach. In the following, there are presented the recent findings under open pond and closed pond culturing techniques for microalgae biomass production.

# **Open pond culture systems**

The oldest, conventional, and commonly used systems for microalgae cultivation are open pond systems. Researchers follow this system mainly due to its maintenance. At the same time, open ponds are mostly preferred because of their lower energy consumption, easy construction procedures, and low operational costs (31,32). The size and shape of these types of ponds are variable; usually depth between 1 and 100 cm, and the area consists of about one to several acres. In terms of pond shape, circular and shallow big ponds are very common nowadays along with thin and multilayer configurations to increase the production efficiency (33-35). Although cost is a factor, that's why scientists designed the size and shape of the pond based on algal species and geographical conditions (36). There are a lot of open pond systems available; however, scientist prefers the most common type is the paddle-wheel raceway pond, which has comparatively more advantages than traditional open pond systems. The shape of a typical raceway pond has resembled that of a race track with a paddle wheel. The paddle wheel mixes and circulates the liquid around the entire pond which makes this cultivation system mostly used for microalgae culture at industrial scales (4,31). Not only that, for treating wastewater by microalgae, these ponds looking more perspective than the others (37). According to Marchetti et al., different microalgae species shows the different biochemical composition due to variations in culture and geographical conditions that meant the placement of open pond system also should be considered during pilot-scale productions (38). However, this raceway open pond system also has some limitations. For example, mixing of nutrients, CO2 transfer, and light availability equally is not almost the same (39).

However, although this type of pond has the limitations mentioned above, an open raceway cultivation system is widely used in different parts of the world due to its low energy requirements (32). Moreover, multilayer system incorporation into open pond raceway systems, consisting of a combination of several open tanks placed at different heights, recently makes it more attractive for microalgae cultivation (35,40). Recently, Min et al. (2013) tested a pilot-scale multilayer system of microalgae cultivation, which gave promising results for Chlorella biomass production that would be usable for other types of microalgae (35). Most of the conventional open pond systems face major disadvantages including high land requirements, contamination issues, and CO<sub>2</sub> discharge to the atmosphere, poor light utilization, and continuous evaporation to loss water (39). Therefore, these limitations make the open pond system of microalgae

cultivation less preferable by scientists for research purposes. As microalgae has correlation with its surroundings in natural water bodies, there would be species needed to study specific growth environmental set up for open pond cultivation system prior to start any commercial productions (40). To maintain the research homogeneity, closed photobioreactors (PBR) are preferred for the cultivation of microalgae over the open pond system. Microalgae cultivation using biofilm in liquid suspension is a prospective very recent finding. However, there are many more recent findings have been made, which make this cultivation system promising for future research to make it profitable (Table1).

## **Closed system photobioreactors (PBRs)**

The closed photobioreactors are made of glass or transparent PVC with varying size and shape. They can be located both in outdoors and indoors environmental conditions. The tubular shaped PBRs systems are very popular during microalgae cultivation, although helical, flat panel like and airlift PBRs systems are being improving for different purposes. Not only that, more other shaped PBRs systems has been developed nowadays. Vertical tubular shaped photobioreactors, horizontal tubular photobioreactors, tank photobioreactors and hybrid type PBRs have been developed and trying to improve more for better results (33). However, the tubular PBRs installing horizontally or vertically has been taken in most cases, due to more advantages than limitations and presently considered as more helpful than open pond culture system (42).

However, the Photobioreactor systems seems more advantageous than open pond system, they have some drawbacks also. For example, algae biomass must settled in the bottom if there lack high turbulent flow (4). To overcome this associated problems scientists use pump to lift air within the reactors. Moreover, scientists find out that the outcome of PBRs would be limiting if there arise any design flaws, which can make a large scale productions abortive economically (43,44). Scientists also reported that suitable material selections for construction, relevant shape, size and spacing along with the optimum operational modes could reduce the production costs more than the present (45,46). To overcome the situations related with design flows there need to design effective as much as possible (47). However, these should be very effective based on geographical conditions and microalgae species specific growth environments (48). Chakraborty et al. (2021) investigate the growth rate of several microalgae in PBR system along with different environment conditions set up and found significant findings (49). They reported different microalgae species responses differently in a same given conditions. Besides, report found that the *C. vulgaris* gave much more good results in biomass yield growing in a modified soil extract medium at a pH of 7.2 and 12:12 light: dark conditions (50). However, there are found several reports emphasizing the similar findings for different microalgae species rather than *C. vulgaris* (42,43,51).

Therefore, researchers recently propose hybrid designs, which particularly is being developed combining both tubular and flat panel PBRs systems. Not only had those hybrid systems, Suh and Lee listed some other prospective designs to make it profitable and promising for microalgae cultivation in large scale (52). Recently, the results found in biomass yield and lipid content perspective culturing with helicoidally and horizontal PBRs systems was lucrative for some algae species, but the horizontal PBRs have a lower biomass yield by other report (53). Consequently, the combination of efficient modeling and design of PBRs along with a better understanding of the growth parameters will be helpful to chase the such types of challenges (42,43). Hopefully, outdoor mass cultivation by large PBRs systems along with media alterations to change the biochemical compositions of microalgae would be attractive in future (54,48).

**Table 1.** Some of recent progress in microalgae cultivationsystems (Open pond and PBRs system).

Metho	od Findings	Ref
ОР	1. Using biofilm in liquid suspension	(55)
	2. Alternative media in raceway system	(56)
	3. Using wastewater as nutrient source	(57)
	4. Mixed culture in a same system	(58)
	5. Paddle wheel raceway system	(31)
PBR	1. Outdoor mass cultivation	(54)
	2. Media alterations	(48)
	3. Algal biofilm membrane	(59)
	4. Industrial gas use as byproduct	(60)
	5. New protocol for high yielding	(50)

Industrial flu gas also could reduce the production costs in closed PBRs system that is considering because of its direct environmental benefits (50).

different types open However, ponds and photobioreactors are being commonly used for C. *vulgaris* culturing suitable but photobioreactors comparatively expensive due to its sophisticated design and controlled operations. Recently other scientists found some limitations of this system (28,61). Open pond and PBRs system considered as mostly used cultivation techniques for microalgae biomass production. However, co-culture of microalgae with fungi or other microorganisms also gaining popularity (62). Moreover, co-culture with bacteria are also getting attention for higher biomass yield nowadays (63,64). This co-culture, either fungi or bacteria, provides synergistic effects on microalgae biomass yields (64). In present, the improvement on co-culture methods of microalgae regarded culturing are being as research advancement in microalgae cultivation process, which is being investigated for further development worldwide.

## CONCLUSION

Open pond cultivation systems for microalgae biomass production are very common for large scale demands, although the system has some limitations yet. The PBRs systems are using mainly for pure culture and most predominant culture system for research or laboratory purposes. Both the systems has gain some advancements to avoid their respective limitations recent years, production costs is still in higher, however. Therefore, to make profitable productions more research needed focusing on lowering production costs.

#### **CONFLICT OF INTEREST**

No conflict of interest.

#### REFERENCES

- 1. Tan JS, Lee SY, Chew KW, et al. A review on microalgae cultivation and harvesting, and their biomass extraction processing using ionic liquids. Bioengineered. 2020;11(1):116-129.
- 2. Chaumont D. Biotechnology of algal biomass production: a review of systems for outdoor mass culture. Journal of Applied Phycology. 2004;5:593-60.
- 3. Metting FB. Biodiversity and application of microalgae. Journal of Industrial Microbiology and Biotechnology. 1996;17(5-6):477-89.
- 4. Chisti Y. Biodiesel from microalgae. Biotechnology Advances. 2007;25(3):294-06.
- 5. Hoek C, Hoeck H, Mann D, Jahns H. Algae: an introduction to phycology. Cambridge university press. 1995. pp. 623.
- 6. Martino A, Bartual A, Willis A, et al. Physiological and molecular evidence that environmental changes elicit

morphological inter conversion in the model diatom *Phaeodactylum tricornutum.* Protist. 2011;162:462–81.

- 7. Biller P, Ross AB. Pyrolysis GC–MS as a novel analysis technique to determine the biochemical composition of microalgae. Algal Res. 2014;6:91–7.
- 8. Priyadarshani I, Rath B. Commercial and industrial applications of micro algae—A review. J Algal Biomass Util. 2012;3:89–100.
- 9. Williams PJL, Laurens LML. Microalgae as biodiesel and biomass feedstocks: Review and analysis of the biochemistry, energetics and economics. Energy Environ Sci. 2010;3:554–90.
- 10. Morimoto T, Nagatsu A, Murakami N, Sakakibara J, et al. Anti-tumour promoting glyceroglycolipids from the green alga *Chlorella vulgaris*. Phytochemistry. 1995;40:1433-37.
- 11. Yasukawa K, Akihisa T, Kanno H, Kaminaga T, et al. Inhibitory effects of sterols isolated from *Chlorella vulgaris* on 12-0-tetradecanoylphorbol- 13-acetateinduced inflammation and tumor promotion in mouse skin. Biol Pharm Bull. 1996;19:573-76.
- 12. Singh A, Singh SP, Bamezai R. Inhibitory potential of *Chlorella vulgaris* (E-25) on mouse skin papillomagenesis and xenobiotic detoxication system. Anticancer Res. 1999;19:1887-91.
- Mallick N, Mandal S, Singh AK, Bishai M, Dash A. Green microalga *Chlorella vulgaris* as a potential feedstock for biodiesel. J Chem Technol Biotechnol. 2012;87:137-45.
- 14. Parsaeimehr A, Sun Z, Dou X, Chen YF. Simultaneous improvement in production of microalgal biodiesel and high-value alpha-linolenic acid by a single regulator acetylcholine. Biotechnol Biofuels. 2015;8:1-10.
- 15. Demirbas MF. Biorefineries for biofuel upgrading: A critical review. Appl Energy. 2009;86:151–61.
- Wang B, Li Y, Wu N, Lan C. CO<sub>2</sub> bio-mitigation using microalga. Appl Microb Biotechnol. 2008;79(5):707-18.
- 17. Miao X, Wu Q. Biodiesel production from heterotrophic microalgal oil. Bioresour Technol. 2006;97(6):841-46.
- 18. Sommerfeld MR. Characterization of the growth and lipid content of the diatom *Chaetoceros muelleri*. Journal of Applied Phycology. 1997;9:9-24.
- 19. Bazaes J, Sepulveda C, Acién FG, Morales J, et al. Outdoor pilot-scale production of *Botryococcus braunii* in panel reactors. Journal of Applied Phycology. 2012;24:1353–60.
- 20. Tasić MB, Pinto LFR, Klein BC, Veljković VB, Filho RM. *Botryococcus braunii* for biodiesel production. Renew Sustain Energy Rev. 2016;64:260–70.
- 21. Norsker NH, Barbosa MJ, Vermuë MH, Wijffels RH. Microalgal production—a close look at the economics. Biotechnol Adv. 2011;29:24–7.
- 22. Wen X, Du K, Wang Z, Peng X, Luo L, Tao H, et al. Effective cultivation of microalgae for biofuel production: a pilot-scale evaluation of a novel oleaginous microalga *Graesiella* sp. WBG-1. Biotechnol Biofuels 2016;9:123.
- 23. Qin C, Lei Y, Wu J. Light/dark cycle enhancement and energy consumption of tubular microalgal

photobioreactors with discrete double inclined ribs. Bioresour Bioprocess. 2018;5:28.

- 24. Kothari R, Pandey A, Ahmad S, Kumar A, Pathak VV, Tyagi V. Microalgal cultivation for value-added products: a critical enviro-economical assessment. 3 Biotech. 2017;7:243.
- 25. Olaizola M. Commercial development of microalgal biotechnology: from the test tube to the marketplace. Biomol Eng. 2003;20(4):459-66.
- Wang J, Yang H, Wang F. Mixotrophic cultivation of microalgae for biodiesel production: status and prospects. Appl Biochem Biotechnol. 2014;172:3307-29.
- 27. Cui HW, Meng FP, Li F, Wang YJ. Application of sodium erythorbate to promote the growth of *Chlorella vulgaris.* J Appl Phycol. 2017;3:1135-44.
- 28. Liang Y, Sarkany N, Cui Y. Biomass and lipid productivities of *Chlorella vulgaris* under autotrophic, heterotrophic and mixotrophic growth conditions. Biotechnol Lett. 2009;31:1043-49.
- 29. Yeh KL, Chang JS. Effects of cultivation conditions and media composition on cell growth and lipid productivity of indigenous microalga *Chlorella vulgaris* ESP-31. Bioresour Technol. 2012;105:120-27.
- 30. Najafabadi HA, Malekzadeh M, Jalilian F, Vossoughi M, Pazuki G. Effect of various carbon sources on biomass and lipid production of *Chlorella vulgaris* during nutrient sufficient and nitrogen starvation conditions. Bioresour Technol. 2015;180:311-17.
- 31. Zittelli GC, Biondi N, Rodolfi L, Tredici MR. Photosynthesis in microalgae, in: Richmond A, Hu C (Eds.), Handbook of Microalgal Culture: Applied Phycology and Biotechnology, second ed., Wiley Blackwell. Oxford. 2013, pp. 225–266.
- 32. Fernández FGA, Sevilla JMF, Grima EM. Photobioreactors for the production of microalgae. Rev Environ Sci Biotechnol. 2013;12:131–51.
- Ugwu CU, Aoyagi H, Uchiyama H. Photobioreactors for mass cultivation of algae. Bioresour Technol. 2008; 99:4021–28.
- 34. Apel AC, Pfaffingera CA, Basedahla N, Mittwollena N, et al. Open thin-layer cascade reactors for saline microalgae production evaluated in a physically simulated Mediterranean summer climate. Algal Research. 2017;25:381–90.
- 35. Min M, Hu B, Mohr MJ, Shi A, et al. Swine manurebased pilot-scale algal biomass production system for fuel production and wastewater treatment—a case study. Appl Biochem Biotechnol. 2014;172:1390–06.
- Borowitzka MA. Commercial production of microalgae: Ponds, tanks, tubes and fermenters. J Biotechnol. 1999;70:313–21.
- 37. De Godos I, Mendoza JL, Acién FG, Molina E, Banks CJ, Heaven S. Evaluation of carbon dioxide mass transfer in raceway reactors for microalgae culture using flue gases. Bioresour Technol. 2014;153:307–14.
- 38. Marchetti J, Bougaran G, Jauffrais T, et al. Effects of blue light on the biochemical composition and photosynthetic activity of *Isochrysis sp.* (T-iso). J Appl Phycol. 2013;25:109–19.
- 39. Brennan L, Owende P. Biofuels from microalgae—a review of technologies for production, processing,

and extractions of biofuels and co-products. Renew Sust Energ Rev. 2010;14:557–77.

- 40. Hossen R, Chakraborty S, Karmaker D, Das SK. Physico-chemical parameters and diversity of phytoplankton in Kirtankhola River, Bangladesh. Current World Environment. 2021;16(1):190-97.
- 41. Zhou W, Chen P, Min M, et al. Environment enhancing algal biofuel production using wastewaters. Renew Sust Energ Rev. 214;36:256–69.
- 42. Pham H M, Kwak HS, Hong ME, et al. Delopment of an XShape airlift photobioreactor for increasing algal biomass and biodiesel production. Bioresour Technol. 2017;239:211–8.
- 43. Jiménez GA, Adam MM, Franco NM, Guerrero RG. Greybox model identification of temperature dynamics in a photobioreactor. Chem Eng Res Des. 2017;121:125–33.
- 44. Wang T, Hsu C-L, Huang C-H, Hsieh Y-K, Tan C-S, Wang C-F. Environmental impact of CO<sub>2</sub>-expanded fluid extraction technique in microalgae oil acquisition. J Clean Prod. 2016;137:813–20.
- 45. Huang Q, Jiang F, Wang L, Yang C. Design of photobioreactors for mass cultivation of photosynthetic organisms. Engineering. 2017;3:318–29.
- 46. Sheng A, Bilad M, Osman N, Arahman N. Sequencing batch membrane photobioreactor for real secondary effluent polishing using native microalgae: process performance and full-scale projection. J Clean Prod. 2017;168:708–15.
- 47. Arcigni F, Friso R, Collu M, Venturini M. Harmonized and systematic assessment of microalgae energy potential for biodiesel production. Renew Sustain Energy Rev. 2019;101:614–24.
- 48. Vo HNP, Ngo HH, Guo W, et al. A critical review on designs and applications of microalgae-based photobioreactors for pollutants treatment. Sci Total Environ. 2018.
- 49. Chakraborty S, Karmaker D, Rahman MA, et al. Impacts of pH and salinity on community composition, growth and cell morphology of three freshwater phytoplankton. Plant Science Today. 2021;8(3):655–61.
- 50. Fernández FGA, Fernández JM, Sánchez JA, et al. Airlift-driven external-loop tubular photobioreactors for outdoor production of microalgae: assessment of design and performance. Chem Eng Sci. 2001;56(8):2721–32.
- 51. Ortiz MEY, Casazza AA, Aliakbarian B, et al. Production of *Chlorella vulgaris* as a source of essential fatty acids in a tubular photobioreactor continuously fed with air enriched with  $CO_2$  at different concentrations. Biotechnology Progress. 2014;30:916–22.
- 52. Vaičiulytė S, Padovani G, Kostkevičienė J, Carlozzi P. Batch growth of *Chlorella vulgaris* CCALA 896 versus semi-continuous regimen for enhancing oil-rich biomass productivity. Energies. 2014;7:3840-57.
- 53. Akter R, Hossain SMM, Zhe W, Kermanee P, Juntawong N. Enhanced lipid production in *Dunaliella salina* grown under high light intensity by shifting the culture from high to low nitrogen concentration. Adv Environ Biol. 2016;10:18–29.

- 54. Rodolfi L, Zittelli GC, Bassi N, Padovani G, Biondi N, Bonini G, Tredici MR. Microalgae for oil: Strain selection, induction of lipid synthesis and outdoor mass cultivation in a low-cost photobioreactor. Biotechnol Bioeng. 2009;102:100–12.
- 55. Johnson KR, Admassu W. Mixed algae cultures for low cost environmental compen- sation in cultures grown for lipid production and wastewater remediation. J Chem Technol Biotechnol. 2013;88:992–98.
- 56. Schneider DS, Lima RDC, Hoeltz M, et al. Life cycle assessment of microalgae production in a raceway pond with alternative culture media. Algal Res. 2018; 32:280–92.
- 57. Mu D, Min M, Krohn B, et al. Life cycle environmental impacts of wastewater-based algal biofuels. Environ Sci Technol. 2014;48:11696–704.
- Ji B, Zhang W, Zhang N, et al. Biofilm cultivation of the oleaginous microalgae. Bioprocess Biosyst Eng. 2014; 37:1369–75.
- 59. Gao F, Yang Z-H, Li C, et al. A novel algal biofilm membrane photobioreactor for attached microalgae growth and nutrients removal from secondary effluent. Bioresour Technol. 2015;179:8–12.
- 60. Nayak M, Karemore A, Sen R. Sustainable valorization of flue gas  $CO_2$  and wastewater for the production of microalgal biomass as a biofuel feedstock in closed and open reactor systems. RSC Adv. 2016;6: 91111–120.
- 61. Lee 2001. Microalgal mass culture systems and methods: their limitation and potential. J Appl Phycol. 13:307-15.
- 62. Gonzalez LE, Bashan Y. Growth promotion of the microalgae *Chlorella vulgaris* when coimmobilized and cocultured in alginate beads with the plant growth-promoting bacteria *Azospirillum brasilense*. Appl Environ Microbiol. 2000;66:1537–41.
- 63. Srinuanpan S, Cheirsilp B, Prasertsan P, Kato Y. Photoautotrophic cultivation of oleaginous microalgae and co-pelletization with filamentous fungi for cost effective harvesting process and improved lipid yield. Aquac Int. 2018;26:1493–509.
- 64. Bashan LE, Bashan Y. Joint immobilization of plant growth-promoting bacteria and green microalgae in alginate beads as an experimental model for studying plant-bacterium interactions. Appl Environ Microbiol. 2008;74:6797–802.
- 65. Wrede D, Taha M, Miranda AF, et al. Co-cultivation of fungal and microalgal cells as an efficient system for harvesting microalgal cells, lipid production and wastewater treatment. PLoS ONE. 2014;9(11): e113497.

#### Acknowledgements

The author is thankful to Riyad Hossen, lecturer of the Department of Botany, University of Barishal, Bangladesh for providing some literatures, guidelines and suggestions during manuscript preparation.

## Author's contributions

All three authors contributed equally.

#### To cite the article

Rasel I, Karim S, A mini review on microalgae biomass production: Recent progress in cultivation systems. Plants and Ecosystem. 2021;1:15-20.